

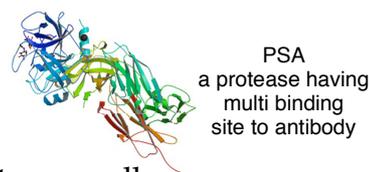
# Plasmonic nanosensors with inverse sensitivity by means of enzyme-guided crystal growth

Laura Rodriguez-Lorenzo, Roberto de la Rica\*, Ramon A. Alvarez-Puebla, Luis M. Liz-Marzan and Molly M. Stevens\* *Nature Materials* 2012, 11, 604–607.

## 1. Introduction

### 1.1. Target molecule: prostate-specific antigen (PSA)

- PSA is a biomarker of prostate cancer (前立腺がん).
- Amount of a biomarker increases due to growth of tumor cells.



=> Detection of a biomarker at low concentration is needed for the early treatment of cancer.

### 1.2. Problem of conventional nanosensors

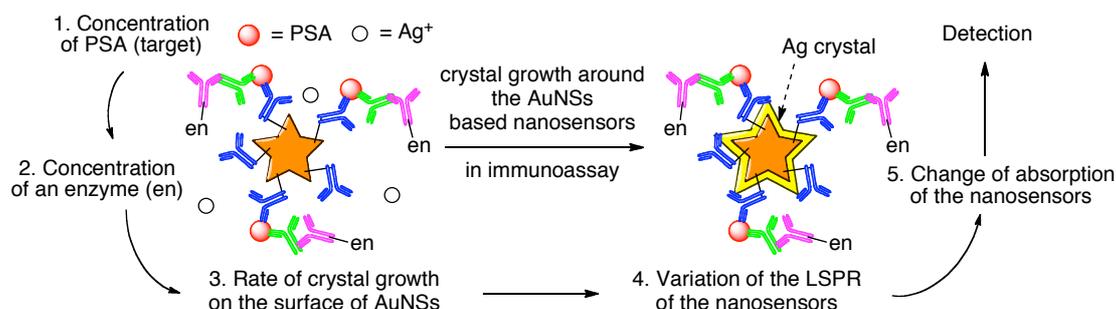
- Conventional nanosensors generate a signal due to directly proportional to the concentration of the target molecule.

=> Difficult to detect with confidence at ultralow concentrations of the molecule

### 1.3. This work

“Nanosensors made of enzyme-modified gold nanostars (AuNSs) as a plasmonic transducer with inverse sensitivity”

- Key point: Inverse sensitivity for the concentration of target molecule
- Strong signal is generated at lower concentration of PSA.
- Information of the concentration of PSA is converted to change of absorption of the AuNSs based nanosensors through enzyme-guided crystallization of silver nanocrystal on the surface of nanosensors (Figure 1).
- Detection of PSA at ultralow concentration
- $10^{-18} \text{ g mL}^{-1}$  ( $= 4 \times 10^{-20} \text{ M}$ ) as a lowest concentration (Previous record: Digital ELISA<sup>1</sup>:  $10^{-17} \text{ g mL}^{-1}$ ) => One order magnitude lower!



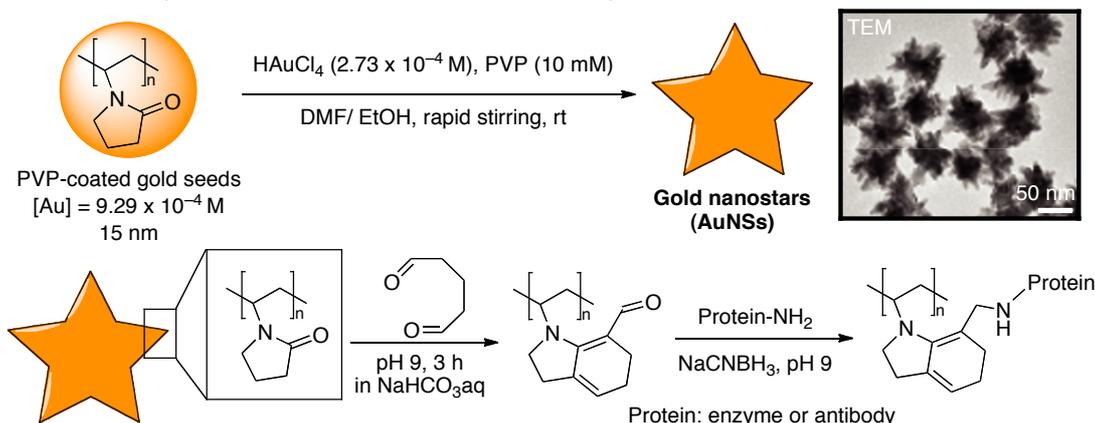
**Figure 1.** Signal translation by the AuNSs based plasmonic nanosensors in immunoassay of PSA, LSPR: localized surface plasmon resonance.

## 2. Results and Discussion

### 2.1. Preparation of protein-modified AuNSs (Scheme 1)

- AuNSs were prepared from poly(vinylpyrrolidone) (PVP)-coated gold seeds.<sup>2</sup>
- Proteins such as enzyme or antibody were combined at the PVP moiety.

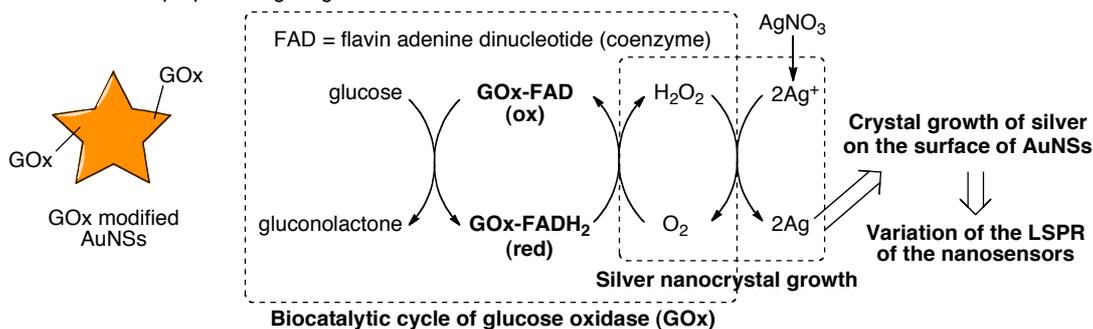
**Scheme 1.** Preparation of AuNSs and covalent attachment of proteins to PVP-stabilized AuNSs.



### 2.2. Signal-generation mechanism by means of enzyme-guided crystal growth (Proof of principle experiment) (Scheme 2)

- AuNSs were modified with glucose oxidase (GOx).
  - Hydrogen peroxide is generated due to reduction of oxygen in biocatalytic cycle of GOx.
  - Silver crystals grow on the surface of AuNSs due to reduction of  $\text{Ag}^+$  by hydrogen peroxide.
- => Information of the concentration of GOx is converted to the rate of crystal growth.
- Localized surface plasmon resonance (LSPR) of the AuNSs changes in proportion to the signal from crystal growth because condition of LSPR changes when silver coats the surface of AuNSs.<sup>3</sup>
  - The signal is amplified in the biocatalytic cycle.

**Scheme 2.** The proposed signal-generation mechanism of GOx-attached AuNSs.



### 2.3. Inverse sensitivity in the plasmonic nanosensors (Figure 2)

- The magnitude of the signal registered by plasmonic nanosensors depends on the rate of crystallization of silver nanocrystal.

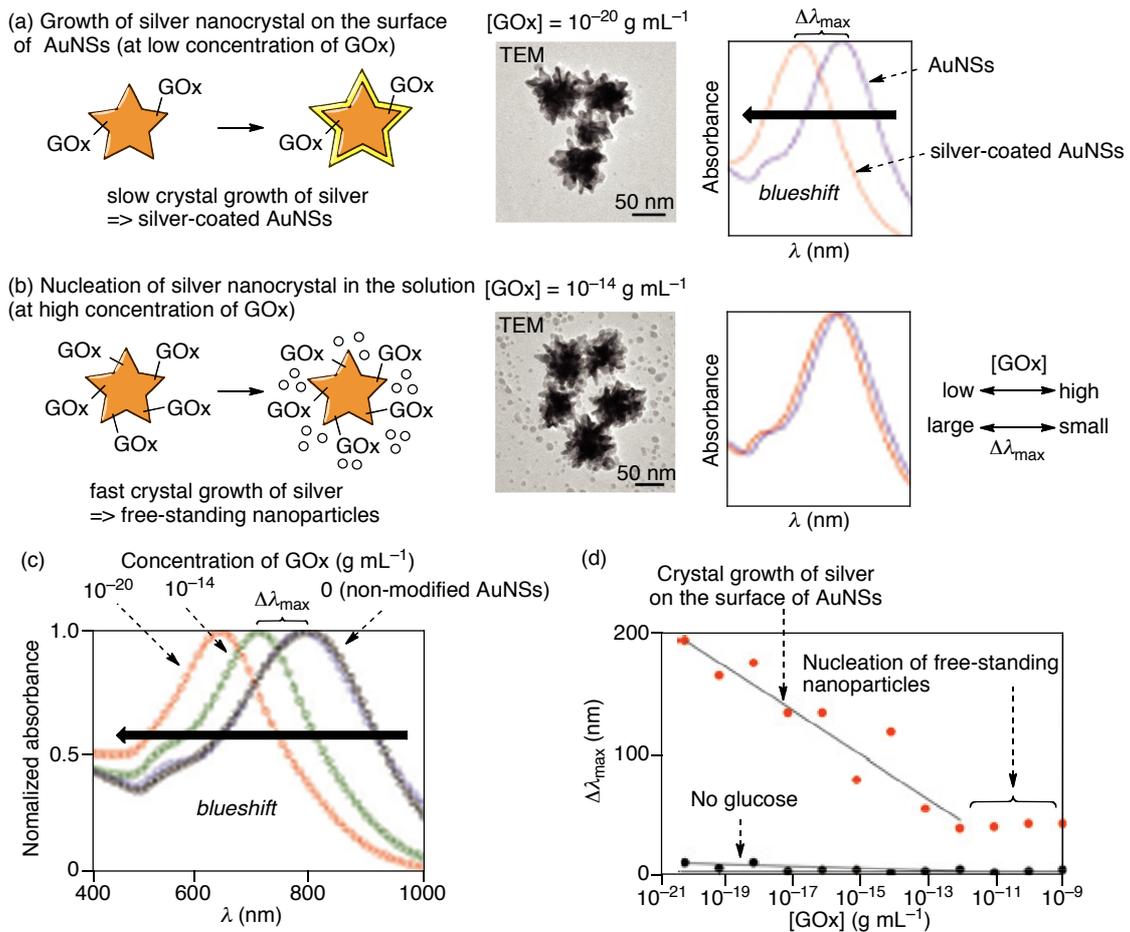
(i) When silver crystals grew **slowly** at low concentration of GOx, the growth of a silver coating on the existing nanocrystal (AuNSs) was favored (Figure 2a).<sup>4</sup>

=> Visible/near-infrared spectra of the nanosensors blueshifted due to variation of the LSPR of the nanosensors (When the [Ag]/[Au] ratio of the plasmonic transducer is higher, the blueshift in the spectra becomes larger.)<sup>3</sup>

(ii) When silver crystals grew **fast** at high concentration of GOx, the nucleation of free-standing small particles was favored (Figure 2b).<sup>4</sup>

=> No change in the visible/near-infrared spectra

- The blueshift of  $\lambda_{\max}$  in the spectra was larger at lower concentration of GOx due to the slow crystallization (Figure 2c and 2d).



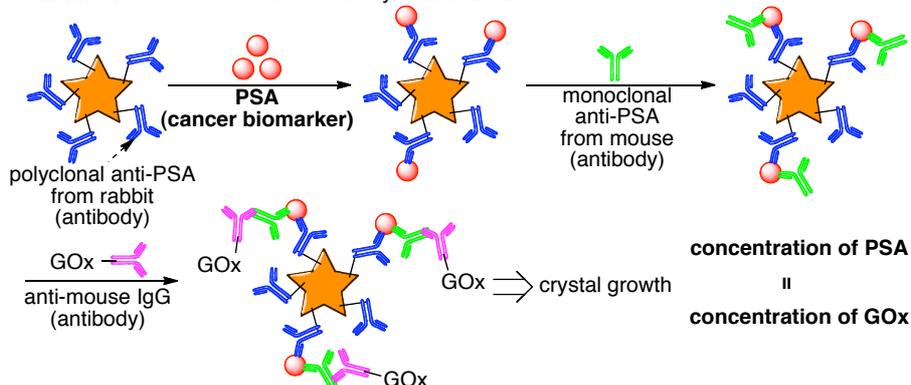
**Figure 2.** Inverse sensitivity in plasmonic nanosensors. (a) Silver nanocrystal growth on the surface of AuNSs at low concentration of GOx, (b) Nucleation of free-standing nanoparticles at high concentration of GOx, (c) Visible/near-infrared spectra of the nanosensors, (d) Relationship between the blueshift of  $\lambda_{\max}$  and the concentration of GOx.

## 2.4. Immunoassay for the detection of PSA with AuNSs based nanosensors

- AuNSs were modified with anti-PSA antibody.
- PSA was captured between the AuNSs and GOx via antibodies (Scheme 3).

=> The concentration of PSA is the same as the concentration of GOx.

**Scheme 3.** PSA detection with antibody-modified AuNSs.

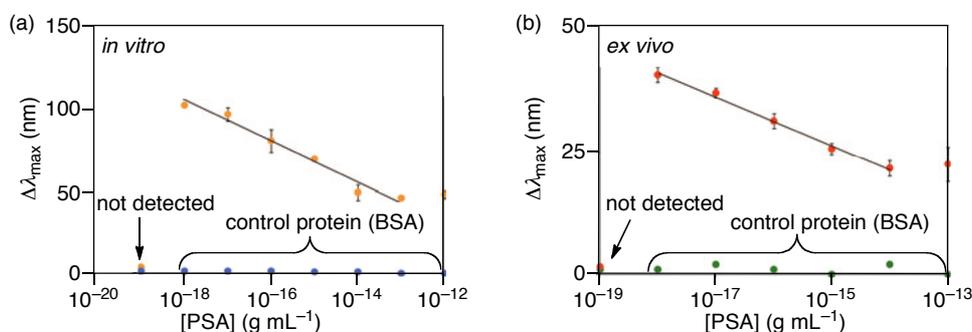


- Ranges of the concentration of PSA detection

– *in vitro*: from  $10^{-13}$  g mL<sup>-1</sup> to  $10^{-18}$  g mL<sup>-1</sup> (Figure 3a)

– *ex vivo* (whole serum): from  $10^{-14}$  g mL<sup>-1</sup> to  $10^{-18}$  g mL<sup>-1</sup> (Figure 3b).

=> Lower limit of PSA detection was  $10^{-18}$  g mL<sup>-1</sup> (=  $4 \times 10^{-20}$  M = 24000 molecules L<sup>-1</sup>).



**Figure 3.** Relationship between the blueshift of  $\lambda_{\max}$  Visible/near-infrared spectra of the AuNSs nanosensors and the concentration of PSA (a) *in vitro* and (b) *ex vivo*, BSA: bovine serum albumin.

## 3. Summary

- The inverse sensitivity phenomenon reported here was possible by controlling the kinetics of crystal growth with enzyme, which in turn determined the signal registered by the plasmonic transducer.

<sup>1</sup> Rissin D. M. et al. *Nature Biotechnol.* **2010**, *28*, 596–599.

<sup>2</sup> Kumar, P. S.; Pastoriza-Santos, I.; Rodriguez-Gonzalez, B.; Garcia de Abajo, F. J.; Liz-Marzan, L. M. *Nanotechnology*, **2007**, *19*, 015606.

<sup>3</sup> Cardinal, M. F.; Rodriguez-Gonzalez, B.; Alvarez-Puebla, R. A.; Perez-Juste, J.; Liz-Marzan, L. M. *J. Phys. Chem. C*, **2010**, *114*, 10417–10423.

<sup>4</sup> Jana, N. R.; Gearheart, L.; Murphy, C. J. *Chem. Mater.* **2001**, *13*, 2313–2322.