Synthesis and Biological Evaluation of QRSTUVWXYZA' Domains of Maitotoxin

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§1. Introduction

1-1. Maitotoxin (MTX)

1976: First isolation from Ctenochaetus striatus (a kind of fish) by T. Yasumoto

1988: Examination of chemical properties by A. Yokoyama, M. Murata, Y. Oshima, T. Iwashita and T. Yasumoto

1996: Determination of complete structure by T. Nonomura, M.Sasaki, N. Matsumori, M. Murata, K. Tachibana,

Y. Kishi and T. Yasumoto



1–2. Hypothetical Bioactivities (**Exact mechanism is still under investigation*)

◆ Sinkins' Hypothesis^[1]...**MTX** converts the Ca²⁺ pump into the Ca²⁺ –permeable nonselective cation channel.

- The structurally related marine toxin, **Palytoxin**, binds to the Na⁺–K⁺–ATPase and converts the Na⁺ pump into a nonselective cation channel (*Figure 2*).
- Ca²⁺–ATPase overexpressed insect cells and human kidneys cells were subjected to **MTX** and showed increase in **MTX**–induced whole cell membrane current.

• Murata's Hypothesis^[2]...W–F' domain works as an anchor that binds to the α -helix active site

- Yessotoxin's Ladder–Shaped Polyether structure, which is similar to W–F' domain of α -helix **MTX**, binds to the α -helix peptide active site of Ca²⁺–ATPase.
- The average distance of ether oxygen atoms on one side matches the α -helix pitch of the active site (*Figure 4*).

? MTX binds to active site of Ca^{2+} -ATPase with W-F' domain and converts it to Ca^{2+} -permeable nonselective cation channel.



Figure 3. YTX, Names of rings correspond to similar rings of MTX.



with Palytoxin

OSO₃Na

Figure 4. Possible structure of **MTX** interacting with active site

1-3. Remaining Challenges and This Work

✗ Total synthesis has not yet been achieved => Fragments coupling entails huge importance to accomplish total synthesis

✗ Mechanism of bioactivity is unknown

=> Q-A' domain contains the fragment promisingly binds to the active site => Understanding bioactivity provides insights into anticancer drug discovery

§2. Results and Discussion (*Some parts are omitted accordingly for clarity and space limitation)

2-1. Synthesis of WXYZA' domain, Synthesis of WXYZA' Ketophosphonate 13

Scheme 1. Synthesis of 1-7, Cascade Takai-Lombardo Olefination/Ring Closing Metathesis and its essence





2-2. Synthesis of QRSTU Aldehyde 18, Fragment Coupling and Completion of QRSTUVWXYZA' Domain

Scheme 4. Synthesis of 14-20, coupling of Q-U domain and W-A' domain by Honor-Wadsworth Emmons Coupling

t-Bu 1) TEMPO (cat.), PhI(OAc) t-Bu MeOBn O MeOBn C Pd/C (cat.), H S MeOBr 2) Ph₃P=CH S U т ... Т R R Ò O 0 R 0 Йe Q OBn н Мe Ňе Q Q OН Me Me Me н ò O. 14 O 15 16 Me Me previously synthesized Me OBn OBn TESO Ĥ o Me \cap 1) TPAP cat. NMO 17: X = O, H 18: X = O Me Me 0 TESC 0 Х w U т s Me z u s Т Me 1) TBAF 0 2) 13 (1 eq) TBSC Α' O R O 2) TESOTf 3) PPTS Ba(OH)2•8H2O Ňе Q R O C Me Йe Q then 18 Me ò O Me THF: H₂O = 6:1 25 °C, 4.5 h 17 19 Me TRDPSC Horner–Wadsworth–Emmons Coupling TESC Me Me \cap х w т S \cap Me z Me TBSC R Α 0 Mel 1) TBAF Me Q [(PPh₃)CuH]₆ Me 2) TESOTf H Ó 20 Me TBDPSC Scheme 5. Synthesis of 21–24, reductive hydroxyketone ring closure and its essence [4] TES Me Me \cap х W U Me w Ζ TES O Z A' R 0 A' Mell MeOBr Мe Q DTES v υ т s 0 Me C R BiBr₃ (0.5 M in MeCN, 3 eq) TESH (50 eq) `O 0 O Me 7.5 Me Q Me 21 22 O TESC MeCN:DCM = 4:1, -10 °C, 2 h HC Me **Reductive Hydroxyketone Ring Closure** Pd(OH)₂/C (cat.), Me₂C(OMe)₂, CSA (cat.) Et₃Si Essence w Ĥ BiBr₃ Ò 0 بحر Me O U Ĥ v ŵ υ H₂O OBr Me Me Х ✓ Stereoselective ring w 22 Н ö BrBiO 21 Me formation A' O MeOBr ν U s 2HBr т Me Et₃SiH TES deprotection, R 0 0 Мe Ĥ Q Et₃SiOSiEt₃ taking advantage of 23 07 нÌ overall yield 31.3% a byproduct HBr Me Et₃SiOH Br Me OH w Ĥ B Me w Me 0 w U ν HO U ν Z HC **A**' O MeOH U т -0 Ъ R 0 Τ̈́Η Ĥ Мe Q H₂O 24 07 н overall yield 37.8% HO Me Et₃SiBr U v HC HB Et₃SiBr Confirmed by NOE and ¹³C NMR

Et₃SiOH

2-3. Biological Evaluation

- 19 different fragments, including previously synthesized A–E, A–G, Q–U, Q–A', W–A' and C'–F' domains and its analogs, were subjected to rat glioma C6 (a kind of cancer) cells (*Figure 5*) and human tumor cells.
- => Compound 24, 25 (Q–A') and 26 (C'–F') (*Figure 6*) gave a positive reaction to the Ca^{2+} influx examination.
- => Compound 24 exhibited significant growth inhibition against 10 different humane tumor cells.
- Others, A-E, A-G, Q-U domains and its analogs were completely inactive or slightly active.





Figure 6. Partial structures of MTX that induced Ca²⁺ influx

- \checkmark Some domains bound to the active site, while all domains did not induce Ca²⁺ influx.
- \Rightarrow Binding domains and Ca²⁺ influx inducing domains are not the same.
- => More than 2 domains play a role of converting Ca^{2+} -ATPase to Ca^{2+} -permeable nonselective cation channel.
- ✔ Q-A' domain and C'-F' domain effectively bound to the active sites (consistent with Murata's hypothesis).
- ✔ W-A' domain nor S-U domain were not active (inconsistent with Murata's Hypothesis).

=> Both S–U domain and W–A' domain are necessary to bind to active sites.

§3. Conclusion

- ✔ Succeeded in synthesizing QRSTUVWXYZA' domains, which was remarkable advance towards total synthesis
- ✔ Evaluated bioactivity of different 19 domains and got new insight into the mechanism of bioactivity of MTX

✔ Promising anticancer activity was observed

§4. References

- [1] W. G. Sinkins et al. Am. J. Physiol. Cell Physiol. 2009, 297, C1533–C1543.
- [2] M. Murata et al. Bull. Chem. Soc. Jpn. 2008, 81, 307–319.
- [3] K. C. Nicolaou et al. J. Am. Chem. Soc. 1989, 111, 5321-5330.
- [4] P. A. Evans et al. J. Am. Chem. Soc. 2003, 125, 11456–11457.

Abbreviations:

MNBA = 2,6-methylnitrobenzoyl anhydride; TPAP = tetra-n-propylammonium perruthenate; NMO = N-methylmorpholine-N-oxide; PPTS = pyridineium p-toluene sulfonate; $CAS = (\pm)$ -camphor-10-sulfonic acid